



A Sysmex Group Company



Instructions For Use

REF: LPH 026-S / LPH026

AML1/ETO (RUNX1/RUNX1T1) Translocation, Dual Fusion Probe



PROFESSIONAL USE ONLY



www.cytoCELL.com

Further information and other languages available at www.ogt.com

Limitations

This device is designed to detect rearrangements with breakpoints in the region covered by the red and green clones in this probe set, which includes the *AML1* and *ETO* (*RUNX1* and *RUNX1T1*) regions. Breakpoints outside this region, or variant rearrangements wholly contained within this region, may not be detected with this product.

The test is not intended for: use as a stand-alone diagnostic, prenatal testing, population-based screening, near-patient testing or self-testing. This product is intended for laboratory professional use only; all results should be interpreted by suitably-qualified staff, taking into account other relevant test results.

This product has not been validated for use on sample types or disease types other than those specified in the intended use.

Reporting and interpretation of FISH results should be consistent with professional standards of practice and should take into consideration other clinical and diagnostic information. This kit is intended as an adjunct to other diagnostic laboratory tests and therapeutic action should not be initiated on the basis of the FISH result alone.

Failure to adhere to the protocol may affect the performance and lead to false positive/negative results.

This kit has not been validated for purposes outside of the intended use stated.

Intended Use

The CytoCell AML1/ETO (RUNX1/RUNX1T1) Translocation, Dual Fusion Probe is a qualitative, non-automated, fluorescence *in situ* hybridisation (FISH) test used to detect chromosomal rearrangements between the 21q22.1 region on chromosome 21 and the 8q21.3 region on chromosome 8 in Carnoy's solution (3:1 methanol/acetic acid) fixed haematologically-derived cell suspensions from patients with confirmed or suspected acute myeloid leukaemia (AML).

Indications

This product is designed as an adjunct to other clinical and histopathological tests in recognised diagnostic and clinical care pathways, where knowledge of *AML1-ETO* (*RUNX1/RUNX1T1*) translocation status would be important for clinical management.

Principles of the Test

Fluorescence *in situ* hybridisation (FISH) is a technique that allows DNA sequences to be detected on metaphase chromosomes or in interphase nuclei from fixed cytogenetic samples. The technique uses DNA probes that hybridise to entire chromosomes or single unique sequences, and serves as a powerful adjunct to G-banded cytogenetic analysis. This technique can now be applied as an essential investigative tool within prenatal, haematological and solid tumour chromosomal analysis. Target DNA, after fixation and denaturation, is available for annealing to a similarly denatured, fluorescently labelled DNA probe, which has a complementary sequence. Following hybridisation, unbound and non-specifically bound DNA probe is removed and the DNA is counterstained for visualisation. Fluorescence microscopy then allows the visualisation of the hybridised probe on the target material.

Probe Information

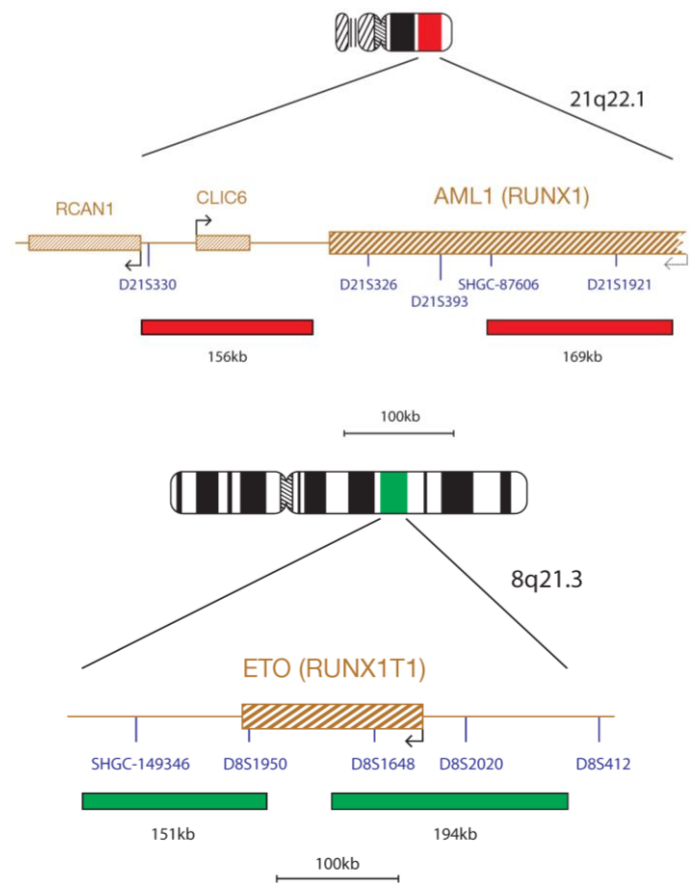
The *RUNX1* (*RUNX1* family transcription factor 1) gene at 21q22.12 is fused with the *RUNX1T1* (*RUNX1* partner transcriptional co-repressor 1) gene at Ensembl location 8q21.3, in the t(8;21)(q22;q22) translocation, found most commonly in patients with acute myeloid leukaemia (AML) FAB (French-American-British classification) type M2.

AML with a *RUNX1-RUNX1T1* fusion resulting from a t(8;21)(q22;q22) translocation is a recognised disease entity according to the World Health Organization (WHO) classification of myeloid neoplasms and acute leukemia¹. The translocation is observed in 10%-22% of patients with AML FAB type M2 and 5%-10% of AML cases overall, most commonly in children and young adults² and is a good prognostic indicator^{3,4,5}. The t(8;21) breakpoint mainly occurs in the intron between exons 5 and 6 just before the transactivation domain and fusion protein created contains the DNA-binding domain of *RUNX1* fused to the transcription factor *RUNX1T1*².

In addition to the reciprocal t(8;21) translocation creating the *RUNX1-RUNX1T1* fusion, variant translocations have also been reported. These variant rearrangements may be cryptic and easily overlooked by G-banding; however, FISH can indicate the presence of such rearrangements².

Probe Specification

AML1, 21q22.12, Red
ETO, 8q21.3, Green



The *AML1* component consists of a 156kb probe, labelled in red, located centromeric to the *AML1* (*RUNX1*) gene that spans the *CLIC6* gene and a 169kb probe covering part of the *AML1* (*RUNX1*) gene, including markers *SHGC-87606* and *D21S1921*. The *ETO* (*RUNX1T1*) component, labelled in green, consists of a 151kb probe covering the centromeric part of the gene and the flanking region and a 194kb probe covering the telomeric part of the gene and the flanking region.

Materials Provided

Probe: 50µl per vial (5 tests) or 100µl per vial (10 tests)

The probes are provided premixed in hybridisation solution (formamide; dextran sulphate; saline-sodium citrate (SSC)) and are ready to use.

Counterstain: 150µl per vial (15 tests)

The counterstain is DAPI antifade (ES: 0.125µg/ml DAPI (4,6-diamidino-2-phenylindole)).

Warnings and Precautions

1. For *in vitro* diagnostic use. For professional use only.
2. Wear gloves when handling DNA probes and DAPI counterstain.
3. Probe mixtures contain formamide, which is a teratogen; do not breathe fumes or allow skin contact. Handle with care; wear gloves and a lab coat.
4. DAPI is a potential carcinogen. Handle with care; wear gloves and a lab coat.
5. Dispose of all hazardous materials according to your institution's guidelines for hazardous waste disposal.
6. Operators must be capable of distinguishing the colours red, blue and green.

- Failure to adhere to the outlined protocol and reagents may affect the performance and lead to false positive/negative results.
- The probe should not be diluted or mixed with other probes.
- Failure to use 10µl of probe during the pre-denaturation stage of the protocol may affect the performance and lead to false positive/negative results.

Storage and Handling



The kit should be stored between -25°C to -15°C in a freezer until the expiry date indicated on the kit label. The probe and counterstain vials must be stored in the dark.



The probe remains stable throughout the freeze-thaw cycles experienced during normal use (where one cycle constitutes the probe's removal from and replacement into the freezer) and is photostable for up to 48 hours after being exposed to continuous lighting conditions. All efforts must be made to limit exposure to light and temperature changes.

Equipment and Materials Necessary but not Supplied

Calibrated equipment must be used:

- Hotplate (with a solid plate and accurate temperature control up to 80°C)
- Calibrated variable volume micropipettes and tips range 1µl - 200µl
- Water bath with accurate temperature control at 37°C and 72°C
- Microcentrifuge tubes (0.5ml)
- Fluorescence microscope (Please see Fluorescence Microscope Recommendation section)
- Phase contrast microscope
- Clean plastic, ceramic or heat-resistant glass Coplin jars
- Forceps
- Calibrated pH meter (or pH indicator strips capable of measuring pH 6.5 – 8.0)
- Humidified container
- Fluorescence grade microscope lens immersion oil
- Bench top centrifuge
- Microscope slides
- 24x24mm coverslips
- Timer
- 37°C incubator
- Rubber solution glue
- Vortex mixer
- Graduated cylinders
- Magnetic stirrer
- Calibrated thermometer

Optional Equipment not Supplied

- Cytogenetic drying chamber

Reagents Needed but not Supplied

- 20x saline-sodium citrate (SSC) Solution
- 100% Ethanol
- Tween-20
- 1M Sodium hydroxide (NaOH)
- 1M Hydrochloric acid (HCl)
- Purified water

Fluorescence Microscope Recommendation

Use a 100-watt mercury lamp or equivalent and oil immersion plan apochromat objectives 60/63x or 100x for optimal visualisation. The fluorophores used in this probe set will excite and emit at the following wavelengths:

Fluorophore	Excitation _{max} [nm]	Emission _{max} [nm]
Green	495	521
Red	596	615

Ensure appropriate excitation and emission filters that cover the wavelengths listed above are fitted to the microscope. Use a triple bandpass DAPI/green spectrum/red spectrum filter or a dual bandpass green spectrum/red spectrum filter for optimal simultaneous visualisation of the green and red fluorophores.

Check the fluorescence microscope before use to ensure it is operating correctly. Use immersion oil that is suitable for fluorescence microscopy and formulated for low autofluorescence. Avoid mixing DAPI antifade with microscope immersion oil as this will obscure signals. Follow manufacturers' recommendations in regards to the life of the lamp and the age of the filters.

Sample Preparation

The kit is designed for use on haematologically-derived cell suspensions fixed in Carnoy's solution (3:1 methanol/acetic acid) fixative, that are prepared according to the laboratory or institution guidelines. Prepare air dried samples on microscope slides according to standard cytogenetic procedures. The AGT *Cytogenetics Laboratory Manual* contains recommendations for specimen collection, culturing, harvesting and for slide making⁶.

Solution Preparation

Ethanol Solutions

Dilute 100% ethanol with purified water using the following ratios and mix thoroughly.

- 70% Ethanol - 7 parts 100% ethanol to 3 parts purified water
- 85% Ethanol - 8.5 parts 100% ethanol to 1.5 parts purified water

Store the solutions for up to 6 months at room temperature in an airtight container.

2xSSC Solution

Dilute 1 part 20xSSC Solution with 9 parts purified water and mix thoroughly. Check pH and adjust to pH 7.0 using NaOH or HCl as required. Store the solution for up to 4 weeks at room temperature in an airtight container.

0.4xSSC Solution

Dilute 1 part 20xSSC Solution with 49 parts purified water and mix thoroughly. Check pH and adjust to pH 7.0 using NaOH or HCl as required. Store the solution for up to 4 weeks at room temperature in an airtight container.

2xSSC, 0.05% Tween-20 Solution

Dilute 1 part 20xSSC Solution with 9 parts purified water. Add 5µl of Tween-20 per 10ml and mix thoroughly. Check pH and adjust to pH 7.0 using NaOH or HCl as required. Store the solution for up to 4 weeks at room temperature in an airtight container.

FISH Protocol

(Note: Ensure that exposure of the probe and counterstain to laboratory lights is limited at all times).

Slide Preparation

- Spot the cell sample onto a glass microscope slide. Allow to dry.
- Immerse the slide in 2xSSC for 2 minutes at room temperature (RT) without agitation.
- Dehydrate in an ethanol series (70%, 85% and 100%), each for 2 minutes at RT.
- Allow to dry.

Pre-Denaturation

- Remove the probe from the freezer and allow it to warm to room temperature (RT). Briefly centrifuge tubes before use.
- Ensure that the probe solution is uniformly mixed with a pipette.
- Remove 10µl of probe per test, and transfer it to a microcentrifuge tube. Quickly return the remaining probe to the freezer.
- Place the probe and the sample slide to prewarm on a 37°C (+/- 1°C) hotplate for 5 minutes.
- Spot 10µl of probe mixture onto the cell sample and carefully apply a coverslip. Seal with rubber solution glue and allow the glue to dry completely.

Denaturation

- Denature the sample and probe simultaneously by heating the slide on a hotplate at 75°C (+/- 1°C) for 2 minutes.

Hybridisation

- Place the slide in a humid, lightproof container at 37°C (+/- 1°C) overnight.

Post-Hybridisation Washes

- Remove the DAPI from the freezer and allow it to warm to room temperature (RT).
- Remove the coverslip and all traces of glue carefully.
- Immerse the slide in 0.4xSSC (pH 7.0) at 72°C (+/- 1°C) for 2 minutes without agitation.
- Drain the slide and immerse it in 2xSSC, 0.05% Tween-20 at RT (pH 7.0) for 30 seconds without agitation.
- Drain the slide and apply 10µl of DAPI antifade onto each sample.
- Cover with a coverslip, remove any bubbles and allow the colour to develop in the dark for 10 minutes.
- View with a fluorescence microscope. (See **Fluorescence Microscope Recommendation**.)

Stability of Finished Slides

Finished slides remain analysable for up to 1 month if stored in the dark at/or below RT.

Procedural Recommendations

- Baking or ageing of slides may reduce signal fluorescence
- Hybridisation conditions may be adversely affected by the use of reagents other than those provided or recommended by Cytocell Ltd
- Use a calibrated thermometer for measuring temperatures of solutions, waterbaths and incubators as these temperatures are critical for optimum product performance.
- The wash concentrations, pH and temperatures are important as low stringency can result in non-specific binding of the probe and too high stringency can result in a lack of signal
- Incomplete denaturation can result in lack of signal and over denaturation can also result in non-specific binding
- Over hybridisation can result in additional or unexpected signals
- Users should optimise the protocol for their own samples prior to using the test for diagnostic purposes
- Suboptimal conditions may result in non-specific binding that may be misinterpreted as a probe signal

Interpretation of Results

Assessing Slide Quality

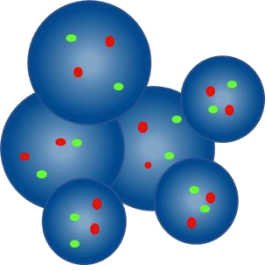
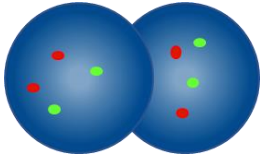
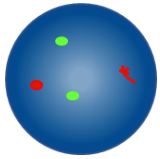
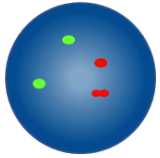
The slide should not be analysed if:

- Signals are too weak to analyse in single filters - in order to proceed with analysis, signals should appear bright, distinct and easily evaluable
- There are high numbers of clumped/overlapping cells obstructing the analysis
- >50% of the cells are not hybridised
- There is excess of fluorescent particles between cells and/or a fluorescent haze that interferes with the signals - in optimal slides the background should appear dark or black and clean

- Cell nucleus borders cannot be distinguished and are not intact

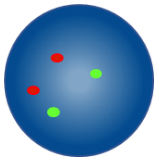
Analysis Guidelines

- Two analysts should analyse and interpret each sample. Any discrepancies should be resolved by assessment by a third analyst
- Each analyst should be suitably qualified according to recognised national standards
- Each analyst should score independently 100 nuclei for each sample. The first analyst should start the analysis from the left side of the slide and the second analyst from the right one
- Each analyst should document their results in separate sheets
- Analyse only intact nuclei, not overlapped or crowded nuclei or nuclei covered by cytoplasmic debris or high degree of autofluorescence
- Avoid areas where there is excess of cytoplasmic debris or non-specific hybridisation
- Signal intensity may vary, even with a single nucleus. In such cases, use single filters and/or adjust the focal plane
- In suboptimal conditions signals may appear diffuse. If two signals of the same colour touch each other, or the distance between them is no greater than two signal widths, or when there is a faint strand connecting the two signals, count as one signal
- If in doubt about whether a cell is analysable or not, then do not analyse it

Analysis Guidelines	
	Do not count – nuclei are too close together to determine boundaries
	Do not count overlapping nuclei – all areas of both nuclei are not visible
	Count as two red signals and two green signals – one of the two red signals is diffuse
	Count as two red signals and two green signals – the gap in one red signal is less than two signal widths

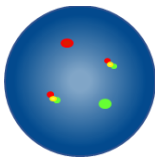
Expected Results

Expected Normal Signal Pattern



In a normal cell, two red and two green signals (2R, 2G) are expected.

Expected Abnormal Signal Pattern



In a cell with a t(8;21)(q21.3;q22.12) translocation, the expected signal pattern will be one red, one green and two fusions (1R, 1G, 2F).

Other signal patterns are possible in aneuploid/unbalanced specimens.

Known Cross-Reactivity

No known cross-reactivity.

Adverse Event Reporting

If you believe this device has malfunctioned or suffered a deterioration in its performance characteristics which may have contributed to an adverse event (e.g. delayed or misdiagnosis, delayed or inappropriate treatment), this must be reported immediately to the manufacturer (email: vigilance@ogt.com).

If applicable, the event should also be reported to your national competent authority. A list of vigilance contact points can be found at <http://ec.europa.eu/growth/sectors/medical-devices/contacts/>.

Specific Performance Characteristics

Analytical Specificity

Analytical specificity is the percentage of signals that hybridise to the correct locus and no other location. The analytical specificity was established by analysing a total of 200 target loci. The analytical specificity was calculated as the number of FISH signals that hybridised to the correct locus divided by the total number of FISH signals hybridised.

Table 1. Analytical Specificity for the AML1/ETO (RUNX1/RUNX1T1) Translocation, Dual Fusion Probe

Probe	Target Locus	No. of Signals Hybridised to the Correct Locus	Total No. of Signals Hybridised	Specificity (%)
Red AML1	21q22	200	200	100
Green ETO	8q21.3	200	200	100

Analytical Sensitivity

Analytical sensitivity is the percentage of scoreable interphase cells with the expected normal signal pattern. The analytical sensitivity was established by analysing interphase cells across different normal samples. The sensitivity was calculated as the percentage of scoreable cells with the expected signal pattern (with a 95% confidence interval).

Table 2. Analytical Sensitivity for the AML1/ETO (RUNX1/RUNX1T1) Translocation, Dual Fusion Probe

No. of Cells with Expected Signal Patterns	No. of Cells with Scoreable Signals	Sensitivity (%)	95% Confidence Interval
4965	5000	99.30	99.03 – 99.50

Characterisation of Normal Cut-off Values

The normal cut-off value, in association with FISH probes, is the maximum percentage of scoreable interphase cells with a specific abnormal signal pattern at which a sample is considered normal for that signal pattern.

The normal cut-off value was established using samples negative for the rearrangement that the probe is intended to detect and the beta inverse function. For each sample, the signal patterns of 100 interphase nuclei were recorded by two independent analysts, totalling 200 per sample.

Table 3. Characterisation of Normal Cut-off Values for the AML1/ETO (RUNX1/RUNX1T1) Translocation, Dual Fusion Probe

Abnormal signal pattern	Number of samples analysed to generate cut-off	Number of nuclei evaluated per sample	Max. no. of false positive signal patterns	Normal cut-off value (%)
1R, 1G, 2F	1290	200	1	2.3

Laboratories must verify cut-off values using their own data^{7,8}.

Reproducibility

Reproducibility was established by three individual laboratories which tested six blinded samples (two negative for the rearrangement, two low positive samples which were 1 to 3 times the cut-off and two high positive samples which contained more than 45% of cells positive for the rearrangement). The analysis was conducted using two replicates of each sample over the course of five non-consecutive days.

All three sites carried out intra-day, inter-day and inter-site testing using the same lot of probe, whilst one of the sites also carried out inter-lot reproducibility using three different lots of probe.

The reproducibility was calculated using the agreement between the variables examined during each test.

Table 4. Reproducibility for the AML1/ETO (RUNX1/RUNX1T1) Translocation, Dual Fusion Probe

Reproducibility study	Sample	Agreement (%)
Intra-day / inter-day / inter-site	Negative	100
	High Positive	100
Inter-lot	Negative	100
	High Positive	100

Clinical Performance

The clinical performance was established using a representative set of unselected patients referred for AML to two different sites (with 100 specimens being collected from site one and 414 being collected from site two). The incident rates of the rearrangements detected by the probe were compared with those gathered from a review of literature sources.

To enable this comparison, the confidence interval indicated by the literature in a population size of 100 samples was calculated by computing 1 – sample proportions test with continuity correction.

Table 5. Clinical Performance for the AML1/ETO (RUNX1/RUNX1T1) Translocation, Dual Fusion Probe

Rearrangement	Prevalence				
	Literature Review (%)	95% LCI (%)	Site 1 (%)	Site 2 (%)	95% UCL (%)
AML with t(8;21)/RUNX1-RUNX1T1 rearrangement	3.8	1.2	0	1.69	10.2

Additional Information

For additional product information please contact the CytoCell Technical Support Department.

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
E: techsupport@cytoCELL.com

W: www.ogt.com

References

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2. Reikvam H, *et al.*, J Biomed Biotechnol. 2011; 2011:104631
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8. Wiktor AE, Dyke DLV, Stupca PJ, Ketterling RP, Thorland EC, Shearer BM, Fink SR, Stockero KJ, Majorowicz JR, Dewald GW. *Preclinical validation of fluorescence in situ hybridization assays for clinical practice*. Genetics in Medicine. 2006;8(1):16-23.

Guide to Symbols

REF	en: Catalogue number
	en: <i>In vitro</i> diagnostic medical device
	en: Batch code
	en: Consult instructions for use
	en: Manufacturer
	en: Use-by date
	en: Temperature limit
	en: Keep away from sunlight
	en: Contains sufficient for <n> tests
	en: Contents

Patents and Trademarks

CytoCell is a registered trademark of CytoCELL Ltd.



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