A Sysmex Group Company

CytoCell

Instructions For Use

REF: LPH 017-S / LPH 017

P53 (TP53) Deletion Probe

CE IVD

PROFESSIONAL USE ONLY

www.cytocell.com

Further information and other languages available at www.ogt.com

Limitations

This device is designed to detect genomic losses larger than the region covered by the red clone in this probe set, which includes the *P53 (TP53)* region. Genomic losses outside this region or partial losses of this region may not be detected with this product.

The test is not intended for: use as a stand-alone diagnostic, prenatal testing, population-based screening, near-patient testing or self-testing. This product is intended for laboratory professional use only; all results should be interpreted by suitably-qualified staff, taking into account other relevant test results.

This product has not been validated for use on sample types or disease types other than those specified in the intended use.

Reporting and interpretation of FISH results should be consistent with professional standards of practice and should take into consideration other clinical and diagnostic information. This kit is intended as an adjunct to other diagnostic laboratory tests and therapeutic action should not be initiated on the basis of the FISH result alone.

Failure to adhere to the protocol may affect the performance and lead to false positive/negative results.

This kit has not been validated for purposes outside of the intended use stated.

Intended Use

The CytoCell *P53 (TP53)* Deletion Probe is a qualitative, non-automated, fluorescence *in situ* hybridisation (FISH) test used to detect chromosomal deletions in the 17p13 region on chromosome 17 in Carnoy's solution (3:1 methanol/acetic acid) fixed haematologically-derived cell suspensions from patients with confirmed or suspected acute myeloid leukaemia (AML), acute lymphoblastic leukaemia (ALL), chronic lymphocytic leukaemia (CLL), multiple myeloma (MM) or non-Hodgkin lymphoma (NHL).

Indications

This product is designed as an adjunct to other clinical and histopathological tests in recognised diagnostic and clinical care pathways, where knowledge of P53 (TP53) deletion status would be important for clinical management.

Principles of the Test

Fluorescence *in situ* hybridisation (FISH) is a technique that allows DNA sequences to be detected on metaphase chromosomes or in interphase nuclei from fixed cytogenetic samples. The technique uses DNA probes that hybridise to entire chromosomes or single unique sequences, and serves as a powerful adjunct to G-banded cytogenetic analysis. This technique can now be applied as an essential investigative tool within prenatal, haematological and solid tumour chromosomal analysis. Target DNA, after fixation and denaturation, is available for annealing to a similarly denatured, fluorescently labelled DNA probe, which has a complementary sequence. Following hybridisation, unbound and non-specifically bound DNA probe is removed and the DNA is counterstained for visualisation. Fluorescence microscopy then allows the visualisation of the hybridised probe on the target material.

Probe Information

The TP53 (*tumor protein p53*) gene at 17p13.1 is a tumour- suppressor gene that has been shown to be deleted in a wide range of human malignancies.

The TP53 gene is one of most important tumour suppressor genes; it acts as a potent transcription factor with fundamental role in the maintenance of genetic stability. Screening for TP53 loss is important as deletions or losses of the short arm of chromosome 17, which includes the TP53 region, are reported in many cancers and are often associated with disease progression, inferior response to treatment and/or a poor prognosis.

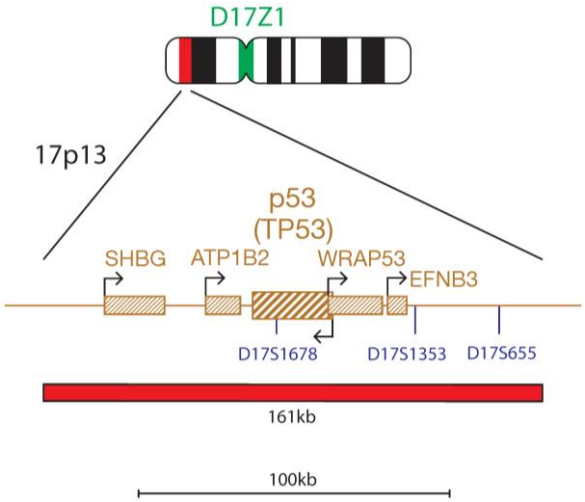
In particular, loss of TP53 is reported in 10% of patients with chronic lymphocytic leukaemia (CLL), and is considered to be the poorest prognostic marker in that disease[1,2]. In acute myeloid leukaemia (AML) and acute lymphoblastic leukaemia (ALL), TP53 loss is associated with a poor outcome and is often seen as a marker of disease progression or secondary disease[3-5].

TP53 loss in patients with multiple myeloma is a late event, where is seen as a marker of disease progression and is associated with a very poor prognosis[6,7].

In non-Hodgkin lymphoma, TP53 losses are reported in diffuse large B- cell lymphoma (DLBCL) often as part of 'dual-hit' lymphoma or plasmablastic phenotypes[8]. In mantle cell lymphoma (MCL), TP53 losses are associated with a poor outcome, and with a dismal outcome when seen with concurrent CDKN2A deletions[9].

Probe Specification

P53, 17p13.1, Red
D17Z1, 17p11.1-q11.1, Green



The P53 probe mix consists of a 161kb probe, labelled in red that covers the whole P53 (TP53) gene and the flanking regions. The probe mix also contains a control probe for the 17 centromere (D17Z1) that is labelled in green.

Materials Provided

Probe: 50µl per vial (5 tests) or 100µl per vial (10 tests)
The probes are provided premixed in hybridisation solution (formamide; dextran sulphate; saline-sodium citrate (SSC)) and are ready to use.

Counterstain: 150µl per vial (15 tests)
The counterstain is DAPI antifade (ES: 0.125µg/ml DAPI (4,6-diamidino-2-phenylindole)).

Warnings and Precautions

1. For *in vitro* diagnostic use. For professional use only.
2. Wear gloves when handling DNA probes and DAPI counterstain.
3. Probe mixtures contain formamide, which is a teratogen; do not breathe fumes or allow skin contact. Handle with care; wear gloves and a lab coat.
4. DAPI is a potential carcinogen. Handle with care; wear gloves and a lab coat.
5. Dispose of all hazardous materials according to your institution's guidelines for hazardous waste disposal.
6. Operators must be capable of distinguishing the colours red, blue and green.
7. Failure to adhere to the outlined protocol and reagents may affect the performance and lead to false positive/negative results.
8. The probe should not be diluted or mixed with other probes.
9. Failure to use 10µl of probe during the pre-denaturation stage of the protocol may affect the performance and lead to false positive/negative results.

Storage and Handling

The kit should be stored between -25ºC to -15ºC in a freezer until the expiry date indicated on the kit label. The probe and counterstain vials must be stored in the dark.

The probe remains stable throughout the freeze-thaw cycles experienced during normal use (where one cycle constitutes the probe's removal from and replacement into the freezer) and is photostable for up to 48 hours after being exposed to continuous lighting conditions. All efforts must be made to limit exposure to light and temperature changes.

Equipment and Materials Necessary but not Supplied
Calibrated equipment must be used:
1. Hotplate (with a solid plate and accurate temperature control up to 80ºC)
2. Calibrated variable volume micropipettes and tips range 1µl - 200µl
3. Water bath with accurate temperature control at 37ºC and 72ºC
4. Microcentrifuge tubes (0.5ml)
5. Fluorescence microscope (Please see Fluorescence Microscope Recommendation section)
6. Phase contrast microscope
7. Clean plastic, ceramic or heat-resistant glass Coplin jars
8. Forceps
9. Calibrated pH meter (or pH indicator strips capable of measuring pH 6.5 – 8.0)
10. Humidified container
11. Fluorescence grade microscope lens immersion oil
12. Bench top centrifuge
13. Microscope slides
14. 24x24mm coverslips
15. Timer
16. 37ºC incubator
17. Rubber solution glue
18. Vortex mixer
19. Graduated cylinders
20. Magnetic stirrer
21. Calibrated thermometer

Optional Equipment not Supplied
1. Cytogenetic drying chamber

Reagents Needed but not Supplied
1. 20x saline-sodium citrate (SSC) Solution
2. 100% Ethanol
3. Tween-20
4. 1M Sodium hydroxide (NaOH)
5. 1M Hydrochloric acid (HCl)
6. Purified water

Fluorescence Microscope Recommendation
Use a 100-watt mercury lamp or equivalent and oil immersion plan apochromat objectives 60/63x or 100x for optimal visualisation. The fluorophores used in this probe set will excite and emit at the following wavelengths:

Fluorophore	$Excitation_{max}$ [nm]	$Emission_{max}$ [nm]
Green	495	521
Red	596	615

Ensure appropriate excitation and emission filters that cover the wavelengths listed above are fitted to the microscope. Use a triple bandpass DAPI/green spectrum/red spectrum filter or a dual bandpass green spectrum/red spectrum filter for optimal simultaneous visualisation of the green and red fluorophores.

Check the fluorescence microscope before use to ensure it is operating correctly. Use immersion oil that is suitable for fluorescence microscopy and formulated for low autofluorescence. Avoid mixing DAPI antifade with microscope immersion oil as this will obscure signals. Follow manufacturers' recommendations in regards to the life of the lamp and the age of the filters.

Sample Preparation
The kit is designed for use on haematologically-derived cell suspensions fixed in Carnoy's solution (3:1 methanol/acetic acid) fixative, that are prepared according to the laboratory or institution guidelines. Prepare air dried samples on microscope slides according to standard cytogenetic procedures. The AGT *Cytogenetics Laboratory Manual* contains recommendations for specimen collection, culturing, harvesting and for slide making[12].

Solution Preparation
Ethanol Solutions
Dilute 100% ethanol with purified water using the following ratios and mix thoroughly.
- 70% Ethanol - 7 parts 100% ethanol to 3 parts purified water
- 85% Ethanol - 8.5 parts 100% ethanol to 1.5 parts purified water

Store the solutions for up to 6 months at room temperature in an airtight container.

2xSSC Solution
Dilute 1 part 20xSSC Solution with 9 parts purified water and mix thoroughly. Check pH and adjust to pH 7.0 using NaOH or HCl as required. Store the solution for up to 4 weeks at room temperature in an airtight container.

0.4xSSC Solution
Dilute 1 part 20xSSC Solution with 49 parts purified water and mix thoroughly. Check pH and adjust to pH 7.0 using NaOH or HCl as required. Store the solution for up to 4 weeks at room temperature in an airtight container.

2xSSC, 0.05% Tween-20 Solution
Dilute 1 part 20xSSC Solution with 9 parts purified water. Add 5µl of Tween-20 per 10ml and mix thoroughly. Check pH and adjust to pH 7.0 using NaOH or HCl as required. Store the solution for up to 4 weeks at room temperature in an airtight container.

FISH Protocol
(Note: Ensure that exposure of the probe and counterstain to laboratory lights is limited at all times).

Slide Preparation
1. Spot the cell sample onto a glass microscope slide. Allow to dry. (**Optional, if using a cytogenetic drying chamber**: slides should be spotted using a cytogenetic drying chamber. The chamber should be operated at approximately 25°C and 50% humidity for optimal cell sample spotting. If a cytogenetic drying chamber is not available, use a fume hood as an alternative).
2. Immerse the slide in 2xSSC for 2 minutes at room temperature (RT) without agitation.
3. Dehydrate in an ethanol series (70%, 85% and 100%), each for 2 minutes at RT.
4. Allow to dry.

Pre-Denaturation
5. Remove the probe from the freezer and allow it to warm to RT. Briefly centrifuge tubes before use.
6. Ensure that the probe solution is uniformly mixed with a pipette.
7. Remove 10µl of probe per test, and transfer it to a microcentrifuge tube. Quickly return the remaining probe to the freezer.
8. Place the probe and the sample slide to prewarm on a 37ºC (+/- 1ºC) hotplate for 5 minutes.
9. Spot 10µl of probe mixture onto the cell sample and carefully apply a coverslip. Seal with rubber solution glue and allow the glue to dry completely.

Denaturation
10. Denature the sample and probe simultaneously by heating the slide on a hotplate at 75ºC (+/- 1ºC) for 2 minutes.

Hybridisation
11. Place the slide in a humid, lightproof container at 37ºC (+/- 1ºC) overnight.

Post-Hybridisation Washes
12. Remove the DAPI from the freezer and allow it to warm to RT.
13. Remove the coverslip and all traces of glue carefully.
14. Immerse the slide in 0.4xSSC (pH 7.0) at 72ºC (+/- 1ºC) for 2 minutes without agitation.
15. Drain the slide and immerse it in 2xSSC, 0.05% Tween-20 at RT (pH 7.0) for 30 seconds without agitation.
16. Drain the slide and apply 10µl of DAPI antifade onto each sample.
17. Cover with a coverslip, remove any bubbles and allow the colour to develop in the dark for 10 minutes.
18. View with a fluorescence microscope. (See **Fluorescence Microscope Recommendation**.)

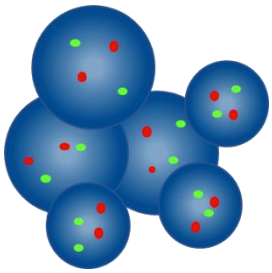
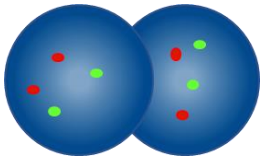
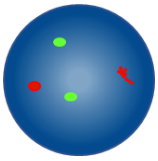
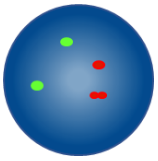
Stability of Finished Slides
Finished slides remain analysable for up to 1 month if stored in the dark at/or below RT.

Procedural Recommendations
1. Baking or ageing of slides may reduce signal fluorescence
2. Hybridisation conditions may be adversely affected by the use of reagents other than those provided or recommended by Cytocell Ltd
3. Use a calibrated thermometer for measuring temperatures of solutions, waterbaths and incubators as these temperatures are critical for optimum product performance.
4. The wash concentrations, pH and temperatures are important as low stringency can result in non-specific binding of the probe and too high stringency can result in a lack of signal
5. Incomplete denaturation can result in lack of signal and over denaturation can also result in non-specific binding
6. Over hybridisation can result in additional or unexpected signals
7. Users should optimise the protocol for their own samples prior to using the test for diagnostic purposes
8. Suboptimal conditions may result in non-specific binding that may be misinterpreted as a probe signal

Interpretation of Results
Assessing Slide Quality
The slide should not be analysed if:
- Signals are too weak to analyse in single filters - in order to proceed with analysis, signals should appear bright, distinct and easily evaluable
- There are high numbers of clumped/overlapping cells obstructing the analysis
- >50% of the cells are not hybridised
- There is excess of fluorescent particles between cells and/or a fluorescent haze that interferes with the signals - in optimal slides the background should appear dark or black and clean
- Cell nucleus borders cannot be distinguished and are not intact

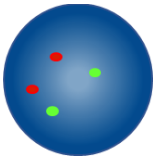
Analysis Guidelines
- Two analysts should analyse and interpret each sample. Any discrepancies should be resolved by assessment by a third analyst
- Each analyst should be suitably qualified according to recognised national standards
- Each analyst should score independently 100 nuclei for each sample. The first analyst should start the analysis from the left side of the slide and the second analyst from the right one
- Each analyst should document their results in separate sheets

- Analyse only intact nuclei, not overlapped or crowded nuclei or nuclei covered by cytoplasmic debris or high degree of autofluorescence
- Avoid areas where there is excess of cytoplasmic debris or non-specific hybridisation
- Signal intensity may vary, even with a single nucleus. In such cases, use single filters and/or adjust the focal plane
- In suboptimal conditions signals may appear diffuse. If two signals of the same colour touch each other, or the distance between them is no greater than two signal widths, or when there is a faint strand connecting the two signals, count as one signal
- If in doubt about whether a cell is analysable or not, then do not analyse it

Analysis Guidelines	
	Do not count – nuclei are too close together to determine boundaries
	Do not count overlapping nuclei – all areas of both nuclei are not visible
	Count as two red signals and two green signals – one of the two red signals is diffuse
	Count as two red signals and two green signals – the gap in one red signal is less than two signal widths

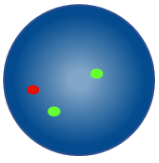
Expected Results

Expected Normal Signal Pattern



In a normal cell, two red and two green signals (2R, 2G) are expected.

Expected Abnormal Signal Pattern



In a cell with a P53 (TP53) deletion the expected signal pattern will be one red and two green signals (1R, 2G).

Other signal patterns are possible in aneuploid/unbalanced specimens.

Known Cross-Reactivity

The green D17Z1 probe may show cross-hybridisation to the centromeres of chromosome 11 and X.

Adverse Event Reporting

If you believe this device has malfunctioned or suffered a deterioration in its performance characteristics which may have contributed to an adverse event (e.g. delayed or misdiagnosis, delayed or inappropriate treatment), this must be reported immediately to the manufacturer (**email:** vigilance@ogt.com).

If applicable, the event should also be reported to your national competent authority. A list of vigilance contact points can be found at: http://ec.europa.eu/growth/sectors/medical-devices/contacts/.

Specific Performance Characteristics

Analytical Specificity

Analytical specificity is the percentage of signals that hybridise to the correct locus and no other location. The analytical specificity was established by analysing a total of 200 target loci. The analytical specificity was calculated as the number of FISH signals that hybridised to the correct locus divided by the total number of FISH signals hybridised.

Table 1. Analytical Specificity for the P53 Deletion Probe

Probe	Target Locus	No. of Signals Hybridised to the Correct Locus	Total No. of Signals Hybridised	Specificity (%)
Red P53	17p13.1	200	200	100
Green D17Z1	17p11.1-q11.1	200	200	100

Analytical Sensitivity

Analytical sensitivity is the percentage of scoreable interphase cells with the expected normal signal pattern. The analytical sensitivity was established by analysing interphase cells across different normal samples. The sensitivity was calculated as the percentage of scoreable cells with the expected signal pattern (with a 95% confidence interval).

Table 2. Analytical Sensitivity for the P53 Deletion Probe

No. of Cells with Expected Signal Patterns	No. of Cells with Scoreable Signals	Sensitivity (%)	95% Confidence Interval
4902	5000	98.04	97.62 – 98.39

Characterisation of Normal Cut-off Values

The normal cut-off value, in association with FISH probes, is the maximum percentage of scoreable interphase cells with a specific abnormal signal pattern at which a sample is considered normal for that signal pattern.

The normal cut-off value was established using samples negative for the rearrangement that the probe is intended to detect and the beta inverse function. For each sample, the signal patterns of 100 interphase nuclei were recorded by two independent analysts, totalling 200 per sample.

Table 3. Characterisation of Normal Cut-off Values for the P53 Deletion Probe

Abnormal signal pattern	Number of samples analysed to generate cut-off	Number of nuclei evaluated per sample	Max. no. of false positive signal patterns	Normal cut-off value (%)
1R, 2G	1600	200	8	6.8

Laboratories must verify cut-off values using their own data[13, 14].

Reproducibility

Reproducibility was established by three individual laboratories which tested six blinded samples (two negative for the rearrangement, two low positive samples which were 1 to 3 times the cut-off and two high positive samples which contained more than 45% of cells positive for the rearrangement). The analysis was conducted using two replicates of each sample over the course of five non-consecutive days.

All three sites carried out intra-day, inter-day and inter-site testing using the same lot of probe, whilst one of the sites also carried out inter-lot reproducibility using three different lots of probe.

The reproducibility was calculated using the agreement between the variables examined during each test.

Table 4. Reproducibility for the P53 Deletion Probe

Reproducibility study	Sample	Agreement (%)
Intra-day / inter-day / inter-site	Negative	95
	High Positive	100
Inter-lot	Negative	100
	High Positive	100

Clinical Performance

The clinical performance was established using a representative set of unselected patients referred for AML or MDS with 100 specimens being collected from the site. The incident rates of the rearrangements detected by the probe were compared with those gathered from a review of literature sources.

To enable this comparison, the confidence interval indicated by the literature in a population size of 100 samples was calculated by computing 1 – sample proportions test with continuity correction.

Table 5. Clinical Performance for the P53 Deletion Probe

Rearrangement	Prevalence			
	Literature Review (%)	95% LCI (%)	Site 1 (%)	95% UCL (%)
AML with *TP53* loss	4.0	1.3	5	10.5
MDS with *TP53* loss	0.6	0.0		5.6

Additional Information
For additional product information please contact the CytoCell Technical Support Department.
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W: www.ogt.com

References

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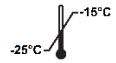
12. Wiktor AE, Dyke DLV, Stupca PJ, Ketterling RP, Thorland EC, Shearer BM, Fink SR, Stockero KJ, Majorowicz JR, Dewald GW. *Preclinical validation of fluorescence in situ hybridization assays for clinical practice*. Genetics in Medicine. 2006;8(1):16–23.

Guide to Symbols

REF	**en:** Catalogue number
	**en:** *In vitro* diagnostic medical device
	**en**: Batch code
	**en:** Consult instructions for use
	**en:** Manufacturer
	**en:** Use-by date
	**en:** Temperature limit
	**en:** Keep away from sunlight
	**en:** Contains sufficient for <n> tests
	**en:** Contents

Patents and Trademarks
CytoCell is a registered trademark of Cytocell Ltd.

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